

ANTIBIOTIC SUSCEPTIBILITY AND RESISTANCE TRENDS IN CITROBACTER FREUNDII AND CITROBACTER KOSERI

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Abstract

Background: *Citrobacter* species, notably *Citrobacter freundii* and *Citrobacter koseri*, are opportunistic pathogens linked to hospital-acquired infections, particularly in immunocompromised patients. These species are known for their ability to produce extended-spectrum beta-lactamases (ESBLs) and carbapenemases, contributing to growing concerns over antibiotic resistance. Understanding their resistance patterns is crucial for effective treatment. This study aimed to assess the antibiotic resistance profiles of *C. freundii* and *C. koseri* isolated from clinical samples. **Materials and Methods:** This prospective observational study was conducted over 1 year and 10 months, involving 150 *Citrobacter* isolates from clinical samples (pus, urine, blood, sputum, and tracheal aspirates) collected in a hospital setting. Isolates were identified using standard biochemical methods, and antimicrobial susceptibility was tested by the Kirby-Bauer disc diffusion method. The presence of ESBL production was assessed using the combination disc method, while carbapenemase production was detected using the Modified Hodge Test (MHT). Data were analyzed using simple percentage calculations. **Results:** Of the 150 isolates, *C. koseri* accounted for 61.33%, and *C. freundii* for 38.67%. Pus samples yielded the highest number of isolates. *C. freundii* exhibited significant resistance to ampicillin, ciprofloxacin, and ceftazidime, while *C. koseri* was more susceptible to most antibiotics, including meropenem and amikacin. ESBL production was observed in 32% of isolates, with *C. freundii* being more prevalent. Additionally, 4.7% of isolates were carbapenemase producers, all identified as *C. freundii*. **Conclusion:** This study highlights the diverse resistance patterns of *C. freundii* and *C. koseri*, particularly the higher resistance of *C. freundii* to several antibiotics. The findings emphasize the importance of continuous surveillance and antibiotic stewardship to manage these infections.

INTRODUCTION

Citrobacter species are abundant in the environment, and have traditionally been considered low-virulence pathogens, producing infections less frequently.^[1] *Citrobacter* species are gram-negative, facultative anaerobic bacilli that are members of the tribe *Citrobacterae* and family *Enterobacteriaceae*. It can be found single or in pairs, and its motile peritrichous flagella usually use citrate as its only carbon source.^[2] Hospital infections can be caused by the bacterium *Citrobacter*. It is frequently associated with brain infections, pneumonia, gastrointestinal infections,

bloodstream infections, and urinary tract infections. Newborns may also be affected, developing joint infections, sepsis, and meningitis.^[3] As per a comprehensive surveillance research carried out in US healthcare facilities, *Citrobacter* infections accounted for 0.8% of Gram-negative infections and 3–6% of all *Enterobacteriales* isolates in hospital settings.^[4]

Among the eleven species—*Citrobacter freundii*, *Citrobacter koseri*, *Citrobacter amalonicus*, *Citrobacter farmeri*, *Citrobacter youngae*, *Citrobacter braakii*, *Citrobacter werkmanii*, *Citrobacter sedlakii*, *Citrobacter rodentium*, *Citrobacter gillenii*, and

Citrobacter murlinae—*Citrobacter koseri* (previously named *C. diversus*), *C. youngae*, *C. braakii*, and *C. amalonaticus* are the most frequently isolated from human clinical specimens.^[5] The two main opportunistic pathogens, *C. koseri* and *C. freundii*, are clearly responsible for the majority of *Citrobacter* infections, with over 80% of patients having underlying medical conditions such as diabetes, cardiovascular disease, renal disease, leukaemia, neurological disease, or urinary tract abnormalities.^[6,7]

Studies on antibiotic resistance in *Citrobacter* isolates have been developing along with the increasing amount of data on hospital infections caused by *Citrobacter* spp., including studies on carbapenemase-producing and Amp C betalactamase strains.^[8]

The aim of this study was to identify the spectrum of the two most prevalent *Citrobacter* species, *Citrobacter freundii* and *Citrobacter koseri*, from clinical samples and their susceptibility pattern. Since these bacteria are commonly associated with hospital infections, especially in immunocompromised patients, understanding their resistance pattern to various antibiotics is essential. The findings from this study will aid clinicians in selecting effective antibiotics and improving patient care.

MATERIALS AND METHODS

The present study was prospective observational study conducted over a period of 1 year and 10 months. All the clinical samples received in the department of microbiology were processed as per standard guidelines. A total of 150 isolates of *Citrobacter* species were identified from various clinical samples. *Citrobacter* spp. was identified by the conventional biochemical methods.^[9] Antimicrobial susceptibility testing was done for all the isolates using Kirby-Bauer disc diffusion method as recommended by Clinical and Laboratory Standards Institute (CLSI) M2-A9.^[10]

ESBL production was detected phenotypically using the combination disc method with ceftazidime/clavulanate and plain ceftazidime discs.

A positive result was confirmed if there was a ≥ 5 mm increase in the zone of inhibition around the ceftazidime/clavulanate disc compared to the plain ceftazidime disc, indicating the presence of ESBL production.^[11]

The Modified Hodge Test (MHT) was performed to detect carbapenemase production. An inoculum of *E. coli* (ATCC 25922), which is susceptible to carbapenems, was streaked on an agar plate. A carbapenem disc (e.g., imipenem) was placed at the center of the plate. The test strain was then streaked in a straight line from the edge of the carbapenem disc to the edge of the plate. After incubation, the plate was examined for the formation of a “cloverleaf” pattern of growth around the streaked area, which would indicate a positive result for carbapenemase production.^[12]

Samples on standard microbiological culture media demonstrating *Citrobacter* species growth were included in the study. Samples with co infections where *Citrobacter* species were not the predominant bacteria and growth from patients receiving antibiotics prior to sample collection or those with incomplete clinical data in requisition form were excluded from the study.

Data was analysed by using simple percentage method.

RESULTS

Out 150 isolates, *C. koseri* (61.33%) was found to be the most common *Citrobacter* species followed by *C. freundii* (38.67%). In the present study, majority of samples yielded *Citrobacter freundii* (84.48%) was found to be from in patients of our hospital. In the present study both species were predominantly observed among male patients (63.3%) followed by female patients (36.7%). In the present study, *C. freundii* was more common in the 31–40 age range, but *C. koseri* typically more common in younger age groups, particularly those between 21 and 40. Both species' overall incidence tends to decline with age, with those over 70 years old seeing the lowest prevalence. [Table 1]

Table 1: Age wise distribution of isolates.

Age in years	<i>C. freundii</i>	<i>C. koseri</i>
1-10	7(12.1%)	5(5.4%)
11-20	1(1.7%)	4(4.3%)
21-30	6(10.3%)	17(18.5%)
31-40	21(36.2%)	38(41.3%)
41-50	9(15.5%)	10(10.9%)
51-60	7(12.1%)	11(12%)
61-70	6(10.3%)	6(6.5%)
>70	1(1.7%)	1(1.1%)
Total	58(100%)	92(100%)

In our study, Both species were frequently isolated from pus samples, with *C. koseri* being slightly more prevalent. *C. freundii* was found in 21 samples (36.2%) and *C. koseri* in 31 samples (33.7%). *C. freundii* was more commonly found in urine samples

compared to *C. koseri*. *C. freundii* was isolated in 17 samples (29.3%) and *C. koseri* in 21 samples (22.8%). *C. freundii* was present in 8 samples (13.8%) and *C. koseri* in 27 samples (29.3%). *C. koseri* was significantly more prevalent in sputum

samples, indicating its stronger association with respiratory infections. [Table 2]

Table 2: Sample wise distribution of isolates

Sample	C. freundii	C.koseri
Pus	21(36.2%)	31(33.7%)
Urine	17(29.3%)	21(22.8%)
Sputum	8(13.8%)	27(29.3%)
Blood	8(13.8%)	10(10.9%)
Tracheal aspirate	2(3.4%)	2(2.2%)
BAL fluid	2(3.4%)	1(1.1%)

In our study, *Citrobacter. koseri* found to be more susceptible to different classes of antibiotics than *C. freundii* overall, especially to Piperacillin/Tazobactam, Meropenem, and Co-Trimoxazole. It was observed that *C. freundii* exhibits higher resistance to Ampicillin, Ciprofloxacin, and Ceftazidime, which is consistent with its higher beta-lactamase production.

All the strains of *C.koseri* exhibited susceptibility to Meropenem. Meropenem and Amikacin are reliable options for both species. Susceptibility pattern was found to be low against various third generation cephalosporins in the study. But cefoxitin, second generation cephalosporin remained as more susceptible to both species. [Table 3]

Table 3: Antibiotic susceptibility of citrobacter species

Antibiotic	C. freundii(n=58)	C.koseri(n=92)
Amikacin	47(81.03%)	81(88.04)
Ampicillin	9(15.5%)	21(22.8)
Piperacillin/Tazobactam	45(77.5%)	90(97.8)
Gentamicin	39(67.2%)	75(81.5)
Ciprofloxacin	22(37.9%)	67(72.8)
Meropenam	51(87.9%)	92(100)
CO-Trimaxazole	38(65.5%)	90(97.8)
Ceftazidime	31(53.4%)	72(78.2)
Ceftriazone	26(44.8%)	57(61.9)
Cefotaxime	29(50%)	42(45.7)
Cefoxitin	45(77.5%)	89(96.8)

In the present study, 48(32%) strains of *Citrobacter* species were found to be ESBL producing. *Citrobacter freundii* (24.67%) was observed as most common ESBL producing strain than *C. koseri* (7.33%). Majority of ESBL producing strains found to be susceptible to Meropenem and Amikacin. Among ESBL producing strains, consistent resistant was observed towards Ampicillin, Ciprofloxacin, and Ceftazidime.

A total of seven strains were found to carbapenamase producing and accounted for 4.7%. All carbapenamase producing strains were *C. freundii*. Carbapenamase producing *Citrobacter* strains were resistant to all cefalosporins, Piperacillin/Tazobactam and Amikacin. All the carbapenamase producing *Citrobacter* strains were resistant to all tested antibiotics in the study.

DISCUSSION

In the present study, a total of 150 *Citrobacter* species were isolated from various clinical samples processed during study period. *Citrobacter koseri* (61.33%) was found more often than *Citrobacter freundii* (38.67%) in the present study, in addition younger patients had a greater prevalence. The age group of 31–40 years old had the highest prevalence of *C. koseri* (41.3%), followed by the age group of 21–30 years old (18.5%), indicating a preference for younger individuals. On the other hand, *C. freundii*

was most prevalent from people between the ages of 31 and 40 (36.2%), followed by people between the ages of 41 and 50 (15.5%), suggesting that it tends to affects middle-aged people. Patients over 70 years old had the lowest prevalence for both species, which is consistent with studies showing a decrease in the frequency of *Citrobacter* infections in aged population.⁵ Numerous variables, including host immunity, comorbidities, and exposure to hospital environments, may have an impact on the age distribution pattern of *C. freundii* and *C. koseri*. The opportunistic pathogen *C. freundii* is frequently identified from patients who have underlying illnesses such as diabetes, renal disease, and cancers, which are more common in middle-aged people.⁶ Furthermore, effective clinical care and antibiotic selection for *Citrobacter* infections can be facilitated by an awareness of age-related susceptibility patterns.^[4]

In the present study, majority of isolates were from pus samples (34.7%). In the study by Dhanya A and Sevitha Bhat, *Citrobacter* species were predominantly isolated from pus samples, accounting for 47.2% of the isolates.¹³ This finding is in line with the present study, where pus samples constituted 36.2% of *C. freundii* and 33.7% of *C. koseri* isolates. These studies collectively highlight the significance of *Citrobacter* species in wound infections. Second most common sample yielded the growth was urine samples(25.3%). This is in agreement with the study

conducted by Metri et al. (24.3%).² On the other hand, a large number of research studies conducted globally have demonstrated that the majority of *Citrobacter* species isolates are found in urine samples.^[2,14,15]

In the present study, susceptibility pattern of *Citrobacter freundii* and *Citrobacter koseri* are two significant bacteria linked to various clinical infections in hospital and community. It was observed that *C. koseri* was more susceptible to a wider range of antibiotics, *C. freundii* exhibited significant resistance to various antibiotics particularly to ampicillin, ciprofloxacin, and ceftriaxone. Both *C. freundii* and *C. koseri* showed a high level of meropenem susceptibility, with *C. koseri* being 100% sensitive. This result is consistent with earlier research showing that carbapenems are effective against *Citrobacter* species because of their broad-spectrum action and capacity to evade several typical resistance mechanisms, including the generation of beta-lactamases.^[16]

In our study, overall ESBL production among *Citrobacter* species was 32%. The prevalence of ESBL production is extremely high compared to the previous study by Kanamori et al. As per the results of Kanamori et al., ESBL production among *Citrobacter* species was 19.3%.^[17] In our research, we found that *C. freundii* had a higher prevalence of ESBL-producing strains than *C. koseri*. This is consistent with other studies that found ESBL production in a significant percentage (20% to 60%) of *C. freundii* isolates.^[16,18] In contrast study by Kanamori et al. showed *C. koseri* as common producer of ESBL production.^[17]

In our study carbapenamase producing strains were 4.7%. All the strains were *Citrobacter freundii*. According to studies, the frequency of *C. freundii* strains that produce carbapenamase varies greatly by the geographical however in some hospital settings, it has been detected in as many as 10–30% of clinical isolates.^[19,20] *C. koseri* is typically less frequently linked to the development of carbapenamase than *C. freundii*. According to certain studies, the prevalence of carbapenamase production in *C. koseri* is very low, ranging from 3% to 15%.^[16,21]

The present comprises of following limitations, *Citrobacter freundii* and *Citrobacter koseri* were the two species of *Citrobacter* that were the subject of the study. Because of its narrow focus, the study does not fully depict the variety of *Citrobacter* species that might be involved in clinical infections. Furthermore, only traditional biochemical procedures were employed for identification, which could have resulted in the omission of certain strains that could have been recognised through the use of more sophisticated molecular methods.

CONCLUSION

This study highlights the antibiogram of *Citrobacter freundii* and *Citrobacter koseri*, with pus samples

yielding the highest number of isolates. *C. koseri* demonstrated higher susceptibility to the tested antibiotics particularly towards piperacillin/tazobactam, meropenem, and cotrimoxazole, whereas *C. freundii* exhibited greater resistance, towards the tested antibiotics. Notably, all carbapenamase-producing strains were *C. freundii*, the findings emphasize the need for continuous surveillance, antibiotic stewardship, and targeted antimicrobial therapy to effectively manage *Citrobacter* infections and mitigate the spread of resistant strains.

REFERENCES

1. Lee R, Choi S-M, Jo SJ, Lee J, Cho S-Y, Kim S-H, et al. Clinical Characteristics and Antimicrobial Susceptibility Trends in *Citrobacter* Bacteremia: An 11-Year Single-Center Experience. *Infect Chemother.* 2019;51:1–9. doi: 10.3947/ic.2019.51.1.1.
2. Metri BC, Jyothi P. Antibiotic sensitivity pattern of *Citrobacter* spp. Isolated from patients with urinary tract infections in tertiary care hospital in South India. *Int J Pharm Pharmaceut Sci.* 2015; 7(1):14–16
3. Ribeiro T. G., Izdebski R., Urbanowicz Paweł and Carmeli Y., Gniadkowski M., Peixe L., Urbanowicz P., et al. (2021). *Citrobacter telavivum* sp. nov. with chromosomal mcr-9 from hospitalized patients. *Eur. J. Clin. Microbiol. Infect. Dis.* 40, 123–131. doi: 10.1007/s10096-020-04003-6.
4. Negrete-González C, Turrubiarres-Martínez E, Briano-Macias M, Noyola D, Pérez-González LF, González-Amaro R, et al. Plasmid carrying blaCTX-M-15, blaPER-1, and blaTEM-1 genes in *Citrobacter* spp. from regional hospital in Mexico. *Infect Dis(Auckl).* 2022;15:11786337211065750. Doi: 10.1177/11786337211065750.
5. Liu L, Wang N, Wu AY, Lin C, Lee C, et al. *Citrobacter freundii* bacteremia: Risk factors of mortality and prevalence of resistance genes. *J Microbiol Immunol Infect* 2018; 51(4):565–572.
6. Ramachandran K., Patel Y., Shetty A. P., Shanmuganathan R. (2022). *Citrobacter koseri* as a rare cause of hematogenous pyogenic spondylodiscitis in young adult – A case report. *J. Orthop. Rep.* 1, 51–54. doi: 10.1016/j.jorep.2022.03.005.
7. Khan S., Taj R., Rehman N., Ullah A., Khan I., Rahman S. (2020). Incidence and antibiogram of β Lactamases-producing *Citrobacter freundii* recovered from clinical isolates in Peshawar, Pakistan. *Pak. J. Zool.* 52, 1877–1882. doi: 10.17582/journal.pjz/20181118151126.
8. Iredell J, Brown J, Tagg K. Antibiotic resistance in Enterobacteriaceae: Mechanisms and clinical implications. *BMJ.* 2016;352:h6420. doi: 10.1136/bmj.h6420.
9. Collee JG, Fraser AG, Marmion BP, Simmons A Collee JG, Miles RS, Watt B. Tests for the identification of bacteria Mackey and McCartney Practical Medical Microbiology. 2006 14th ed. New Delhi, India Elsevier: 131–49.
10. Clinical and Laboratory Standards Institute. Performance Standards for Antimicrobial Susceptibility Testing. Fifteenth Informational Supplement. CLSI Document M100-S15. 2005 Wayne, PA Clinical and Laboratory Standards Institute.
11. Jarlier V, et al. Detection of extended-spectrum beta-lactamase production by a combination disk test. *J Clin Microbiol.* 1988;26(10):2185–2187
12. Lee K, et al. Modified Hodge test for the detection of carbapenamase-producing bacteria: evaluation of a phenotypic test for surveillance. *J Clin Microbiol.* 2001;39(9):3582–3587.
13. Dhanya A, Sevitha Bhat. Clinicomicrobiological study of infections due to *Citrobacter* species. *J Evol Med Dent Sci.* 2015; 4(42):7327–7331
14. Hawaldar R., Sadhna S. Prevalence and drug resistance pattern of *Citrobacter* spp – A retrospective study. *Indian J. Microbiol. Res.* 2019; 6, 142–145. doi: 10.18231/j.ijmr.2019.030

15. Tadesse S., Mulu W., Genet C., Kibret M., Belete M. A. Emergence of high prevalence of extended-spectrum beta-lactamase and carbapenemase-producing enterobacteriaceae species among patients in Northwestern Ethiopia Region. *BioMed. Res. Int.* 2022: 1–9. doi: 10.1155/2022/5727638.
16. Borer, A., & Gilad, J. *Citrobacter* species: A review of the microbiological features, clinical relevance, and management strategies. *Clinical Microbiology Reviews.*2012: 25(4), 784-794.
17. Kanamori H, Yano H, Hirakata Y, Endo S, Arai K, Ogawa M, Shimojima M, Aoyagi T, Hatta M, Yamada M, Nishimaki K, Kitagawa M, Kunishima H, Kaku M. High prevalence of extended-spectrum β -lactamases and qnr determinants in *Citrobacter* species from Japan: dissemination of CTX-M-2. *J Antimicrob Chemother.* 2011 Oct;66(10):2255-62. doi: 10.1093/jac/dkr283. Epub 2011 Jul 6. PMID: 21733965.
18. Zhang, Y., et al. Prevalence of ESBL-producing *Citrobacter* species in a Chinese hospital: A 6-year retrospective study. *Infection and Drug Resistance.*2016: 9, 133-139.
19. Wang, Q., et al. Antimicrobial resistance and carbapenemase production in *Citrobacter* species: A hospital-based surveillance study in China. *Journal of Global Antimicrobial Resistance.*2018: 14, 113-118.
20. Toleman, M. A., et al. Carbapenemase-producing *Citrobacter* species: Clinical implications and management strategies. *Journal of Antimicrobial Chemotherapy.*2014: 69(9), 2263-2270.
21. Jacob, M. A., et al. Prevalence of carbapenemase-producing *Citrobacter* species in a hospital-based study: An emerging problem in nosocomial infections. *Journal of Clinical Microbiology.*2016 54(3), 689-696