

The Role of Aquaporins in Sinonasal Mucosa Physiopathology

Yücel Kurt¹

REVİEW

¹ Finike Government Hospital, Sahilkent, 07740 Finike/Antalya TURKEY

Abstract: The nose, an organ with crucial respiratory and olfactory functions, is the first organ to be exposed to external stimuli. As the first point of contact during inhalation, the sinonasal airway has various functions, including thermoregulation, humidification, removal of airborne particles and response to infectious agents. The sinonasal airway is exposed to many detrimental stimuli, some of which might be allergic. Allergic rhinitis (AR) and chronic rhinosinusitis (CRS) are the most prevalent diseases associated with the respiratory function of the nose. As typical pathological mechanisms, the pathogenesis of both diseases includes many common characteristics, such as edema and hypersecretion of the nasal mucosa. In this process, the roles of water channels, such as aquaporins (AQP), are vital. AQPs are water-specific membrane channel proteins that regulate cellular water homeostasis. Various AQPs are expressed in both nasal respiratory mucosa and olfactory mucosa. The human normal nasal respiratory epithelium contains AQP1, AQP2, AQP3, AQP4, AQP5, AQP7 and AQP111. The expression of AQPs is different in inflammatory diseases, such as AR or CRS than that in normal tissues. Nasal polyp tissue expressed abundant AQP1 than normal tissue, whereas AQP5 level was at lower levels in the sinonasal mucosa in AR and CRS with the polyp. The expression of AQPs in the olfactory epithelium differs from the expression in the respiratory epithelium. In this study, we investigated the relationship between AQPs and the sinonasal epithelium, which reveals a comprehensive overview of the physiopathological connections between allergic rhinitis, rhinosinusitis, and the olfactory system. This study can provide valuable insights into further investigation of AQPs as a potential therapeutic target.

INTRODUCTION

The sinonasal airway acts as a gateway between the external environment and the human body. As the first point of contact during inhalation, the sinonasal airway serves many roles, such as thermoregulation, humidification, removal of airborne particles, and response to infectious agents¹. The sinonasal airway is exposed to a series of harmful stimuli, some of which might be allergic. It may lead to an immunoglobulin-mediated inflammatory response characterized by several symptoms, runny nose, itching, sneezing and nasal congestion. This allergic response to inhaled including stimuli is known as allergic rhinitis and is one of the most prevalent illnesses². The nasal epithelium plays a key role in maintaining the water homeostasis of the airway. The nasal epithelium actively secretes most of the mucus and determines the electrolyte composition of nasal secretions. Nasal hypersecretion is one of the common characteristics of disorders, such as rhinitis and sinusitis ^{1,2}. This determines the electrolyte composition of nasal secretions, covers the airway fluid layer, which surface. The main role of this fluid is to protect the airway epithelium from inhaled hazardous substances. Hence, the nasal respiratory epithelium forms a crucial physiological defense barrier in the body 2,3 . Aquaporins (AQP), also termed water channels, are integral membrane proteins that belong to a large family of major proteins that form pores in biological membranes, facilitate the intercellular transport of water, and regulate water flow osmotically ^{4,5}. AQPs are generally known to be passive carriers of water. Some of these proteins mediate the transport of small solutes, including glycerol⁶. AQPs, which play a role in the transport of water to various cells, passively transport water across cell membranes and prevent the passage of ions and other solutes ^{4,6}. In addition to being critical for water homeostasis, AQPs also act as signaling factors in the physiological transport of molecules regulation of surface expression of other membrane proteins, regulation of cell other than water, adhesion, and cell volume ^{5,7,8}.

This review aims to investigate the relationship between AQPs and the sinonasal epithelium, reveal a comprehensive overview of the physic pathological connections between allergic rhinitis, rhinosinusitis and the olfactory system.

Structure and function of aquaporins

Water is necessary for all molecular interactions, which are indispensable for the survival of living beings, and which are required for anabolic and catabolic reactions. Therefore, it is vital to regu-



Received

Accepted Available online

Keywords.

Aquaporins

Nasal Mucosa Allergic Rhinitis

Physiopathology Quality of Life

Chronic Rhinosinusitis

Corresponding Author: Yücel Kurt

Int J Acad Med Pharm, 2022; 4 (2); 72-77

E-mail; yucelkurt00@gmail.com ORCID; 0000-0001-5111-0240 http://dx.doi.org/10.47009/jamp.2022.4.2.16

Received in revised form

01/05/2022

03/01/2022

03/30/2022

06/21/2022

late the transport of water, which is the primary component of living organisms ⁵. AQPs, also known as water channels, are molecules with protein structures that pass through biological membranes. They assist in the rapid transport of water through membranes and are also crucial in regulating cell volume ^{4,5}. Although water passage is bilateral in diffusion, aquaporins allow unilateral passage controlled by the osmotic gradient ⁷. Because its molecular weight defined in the erythrocyte membrane is 28-kDa, AQP was first termed CHIP28 (channel dependent integral protein-28) and later termed AQP1, as it is used today ⁹. These proteins have a very specific molecular structure that resembles an hourglass and consists mainly of helices. They are found in cells of a great variety of species, including mammals, microorganisms, animals, and plants ^{4,7}. Recent studies have revealed that AQPs regulate other processes, such as cell migration, cell proliferation, and cell adhesion, in addition to the previously mentioned functions⁵. Hitherto, 13 aquaporin isoforms have been discovered in mammals, all involved in critical functions in various tissues 5,8

Members of the AQP family are divided into three subgroups based on their permeability properties:

AQPs with water permeability: AQP0, AQP1, AQP2, AQP4, AQP5, AQP6, AQP8.

AQPs that are permeable to both water and small molecules, such as urea and glycerol: AQP3, AQP7, AQP9, AQP10.

Hybrid AQPs: Those with permeability for water (AQP 11, AQP 12) and glycerol (AOP 11) 5,9 .

Tissue-specific distributions of AQPs in the human body are summarized in Table 1. Many AQP isoforms (AQP1, 3, 4, 5,7, 8, 9, and 11) are found in the brain and nervous system, but AQP1 and AQP4 are found at relatively higher rates in the brain ^{8,10}. AQP1 is expressed in epithelial cells of the choroid plexus and plays a role in the formation of cerebrospinal fluid ^{11,12}. AQP1 is also present in primary sensory neurons and is suggested to play a role in pain perception¹ Eight of the AQPs (AQP0, 1, 3, 4, 5, 7, 9, and 11) are found in the eye. These AQPs play many physiological roles, such as maintenance of the cornea and lens, wound healing, tear osmolarity, and maintenance of retinal homeostasis ^{14,15}. It has been reported that AQPs (AQP1, 2, 3, 4, 5, 6, 8, 10, and 11) found in the human ear are involved in immunological functions as well as in the regulation of neuronal signal transmission and cell movement 16.

In the skin, AQPs are distributed in the epidermis (AQP1, 3, 7, and 10), dermis (AQP1, 3, and 5), and hypodermis (AQP7) aid skin hydration, cell proliferation, immunity, and wound healing ^{8,17}. Kidney AQPs (AQP1, 2, 3, 4, 5, 6, 7, 8, and 11) play key roles in both short-term and long-term regulation of water balance ^{8,18}. AQP3 is expressed in the submucosal glands of the lung, while AQP4 and AOP5 are expressed in the airways and alveolar type I cells. AOP1, AQP3, and AQP9 are expressed in erythrocytes. AQP7, an aquaglyceroporin, is expressed in adipocytes, regulating glycerol transport through the cell⁸.

Relationship between AQPs and nasal respiratory epithelium

The nasal cavity is divided into two parts, the respiratory segment and the olfactory segment. The respiratory segment makes up most of both nasal cavities and is covered by ciliated pseudostratified columnar epithelium, also called respiratory epithelium. This epithelium contains mucus-producing Goblet cells. The secretion of goblet cells is supported by mucus and serous glands in the connective tissue located under the epithelium called the lamina propria. The vessels in the lamina propria form thin-walled cavernous sinusoids, also called cavernous bodies ^{19,20}. Nasal epithelial cells play a crucial role in maintaining an even temperature by moistening and lightly lubricating the surface, thanks to water/ion channels like AQPs^{20,21}. The expression of AQPs in the respiratory epithelium of the nasal cavity is summarized in Figure 1.

It has been revealed that the human nasal cavity has AQP1, AQP2, AQP3, AQP4, AQP5, AQP7 and AQP11 under respiratory physiological conditions 22.

Table 1 Tissue-specific distributions of AQPs in the human body

| GROUP | AQUAPORIN | TISSUES WHERE IT IS FOUND |
|----------------------|-----------|--|
| | | |
| Classical aquaporins | 0 | Lens |
| | 1 | Renal tubules and capillaries, choroid plexus, ciliary epithelium, corneal endothelium, pain-processing C fibers, vascular endothelial tumor vessels, and erythrocytes |
| | 2 | Renal collecting duct |
| | 4 | Astrocytes, retinal Muller cells, lacrimal gland, salivary duct, inner ear, olfactory epithelium, gastric parietal cells, airways, renal collecting duct, placenta, muscle, intestinal epithelium, and glioblastomas |
| | 5 | Corneal epithelium, sweat glands, lacrimal glands, salivary glands, airway submucosal glands, alveolar type I cells, and epidermis |
| | 6 | Intercalated intracellular vesicles in the renal collecting duct |
| | 8 | Intestinal epithelium |
| | | |
| Aquaglyceroporins | 3 | Renal collecting duct, epidermis, conjunctiva, corneal epithelium, immune cells, intestinal epithelium, and erythrocytes |
| | 7 | Fat cells, renal proximal tubule, testis, and myocardium |
| | 9 | Hepatocytes, erythrocytes, and possibly certain brain cells |
| | 10 | Intestinal enterochromaffin cells |
| | | |
| Hybrid aquaporins | 11 | Liver, testis, and intracellular membranes in the renal proximal tubule |
| | 12 | Exocrine pancreas |
| | | |

humans, it is especially remarkable in the kidneys. In addition to being a membrane channel that allows rapid water movement funneled by a transmembrane osmotic gradient, AQP1 also has a secondary function as a cyclic nucleotide-gated ion channel ^{20,23}.

It has been reported that AQP1 is distributed in various parts of



Figure 1. Expression of AQPs in normal nasal cavity respiratory epithelithe nasal cavity respiratory epithelium. In a study conducted on rats, it was reported that it was abundant in the venules in the capillaries under the nasopharyngeal epithelium, in the capillaries of the nasal conchas, and the venous sinuses, whereas it was not detected in the surface epithelium of the nasopharynx and nasal conchas ²². Another study showed that AQP1 is expressed in endothelial cells of blood vessels and surrounding connective tissue cells in olfactory and respiratory mucosa²⁴. Meanwhile, in studies on humans, it has been Although the AQP1 protein is distributed in a variety of tissues in shown that it is localized in fibroblasts on the inferior turbinate tissue,

especially in the subepithelial area and endothelial cells of blood approximately 10-30% of adults and 40% of children, and its AQP1 might be involved in water transfer through the blood vessel wall

AQP2 is found in structures called collecting ducts in the kidney, where it plays a key role in maintaining the body's water balance ²⁷. In a study conducted on lower turbinate tissue from humans, AQP2 was localized in the cytoplasm of epithelial cells and acinar cells²⁵. However, to our knowledge, no other studies have been reported to support these findings, so it is considered that AQP2 has limited distribution to the renal collecting duct, and its significance in nasal cavity respiratory function is relatively weak than other AQPs.

AQP3 are protein channels that can mediate the passage of glycerol, urea, and other small solutes in addition to water molecules. AQP3 is a water channel protein that allows rapid and selective water transport through the membrane of the human respiratory epithelium in response to osmotic gradients ²². AQP3 is also found in the skin, lungs, cornea, esophagus, colon, stomach, liver, intervertebral discs and sperm ^{20,25}. Studies have revealed that AQP3 is abundant in the basal cells of the trachea and nasopharyngeal epithelium and basolateral membranes of the surface epithelial cells of the nasal ^{22,24,31}. AQP3 conchas expression has been shown in immunohistochemically studies on human inferior turbinate tissue and normal sinonasal mucosa ^{25,26}.

AOP4 is the most common AOP in the brain, spinal cord and optic nerve. It is expressed at the highest level in astrocytic foot processes. In the brain, it is involved in the production and absorption of CSF, water transport between the blood-brain barrier and the physiopathology of brain edema ²⁸.AQP4 is also expressed in plays a role in regulating AQP5 expression in the nasal epithelium of epithelial cells of many organs in the human body, such as the kidney, intestine, salivary glands, sensory organs and skeletal muscles 28 Studies have revealed the presence of abundant AQP4 in the that AQP5 expression decreased in the AR group than in the control basolateral regions of acinar cells and columnar cells in the nasal mucosal epithelium and the basolateral membranes of columnar cells of the nasopharyngeal epithelium 20,22,26

AQP5 takes part in the formation of saliva, tears and pulmonary secretions. AQP5 facilitates the secretion of fluid in the submucosal glands indicating that the lumen membrane of serous epithelial cells is the rate-limiting barrier against water movement ^{20,22,30}. In studies on the distribution of AQP5 to nasal tissues, it has been reported that it is expressed in abundance in the apical plasma membrane of the intraepithelial glands of the nasal conchas, on the apical surface of the nasal respiratory epithelium, and in the apical membrane of subepithelial glandular cells in the nasopharynx ^{20,22,24,26}

AQP7 is abundantly expressed in both white and brown adipose tissue ³². It has been shown that in nasal tissue, normal human sinonasal mucosa, AQPs are localized in the surface and cytoplasm of glandular epithelial cells ²⁶. However, no studies have clearly reported the role of AQP7 in nasal tissue. In general, AQP7 may be involved in the movement of water and other molecules (glycerol, urea, and other small solutes) between subepithelial connective tissues and epithelial cells 33.

AQP11 is a relatively recently discovered member of the AQP family, and not much is known about its function yet. AQP11 is expressed in the nasal mucosa in humans and mice. However, to our knowledge, its specific functions in the nasal respiratory epithelium have not yet been studied 34,26

In addition to showing the abundant expression of AQPs in the respiratory epithelium of the nasal cavity, it has also been shown that these proteins participate in normal physiological processes, such as humidification of inhaled air, but may contribute to the pathogenesis of nasal congestion and rhinorrhea²⁰.

Aquaporins and Allergic Rhinitis

Allergic rhinitis (AR) is a type of inflammation of the nasal mucosa that occurs when the immune system overreacts to airborne

vessels and the vascular and connective tissue of normal human si- prevalence tends to increase ^{35,36}. Hence, it is considered an important nonasal mucosa^{25,26}. Collectively, these observations suggest that chronic respiratory disease due to its high prevalence and negative effects on quality of life. Various mechanisms of sinonasal epithelial barrier disruption include antigen proteolytic activity, tight junction disruption mediated by inflammatory cytokines, or exacerbation by environmental stimuli^{2,20}. By stimulating allergy and inflammation, elevated IgE acts on mast cells to induce histamine release, which plays a crucial role in AR. Histamine also plays an important role in the secretory response of the submucosal glands in the nose. Besides, histamine causes vasodilation, tissue edema and sneezing ³⁷. Another characteristic feature of AR is glandular hypersecretion. Many studies have clearly revealed that AQPs play a key role in maintaining fluid balance in airways, such as the nasal cavity. Thus, AQPs have also been associated with dysregulated water metabolism in AR²⁰. The AQP best studied in the context of AR is AQP5, a sub-member of the classical aquaporins family. AQP5, a transmembrane water channel protein, has a crucial role in water transport on the apical surface of the alveolar epithelium, the upper airways, and the submucosal epithelium of the nasopharynx ³⁷. It has been revealed that AQP5 is closely associated with serious glands and upper airway pathologies². Most likely, AQP5 is the primary water channel in the human nasal mucosa, where it functions as a key tight junction protein in maintaining mucosal water homeostasis. It is a key molecular player in fluid secretion and a rate-limiting barrier to secretion observed during allergic inflammation ^{20,30,38}. Recent studies have shown that AQP5 performs this task in the mucosa using the cAMP Protein Kinase A pathway and that it is possible to halt it at various stages. For example, it has been shown that the cAMP-PKA/CREB pathway rats ³⁹. NF-kB plays a significant role in the regulation of AR cytokine networks. In a study conducted with the AR model, it was revealed group, and this decrease was suppressed by NF-kB inhibitor treatment ⁰. In another AR cell culture study by Chang et al., it was reported that the NF-kB pathway suppressed AQP5 expression ⁴¹. Histamine, which induces hypersecretion in the nasal mucosa, is one of the most considerable contributors to the pathophysiology of AR. In another study on human nasal epithelial cells, it was demonstrated that histamine reduces the expression of AQP5¹. Chlorpheniramine has the ability to reverse the impacts of histamine. Antihistamines, such as chlorpheniramine, are among the most commonly prescribed medications for AR. Chang et al. showed that a significant dosedependent increase occurred in the expression of membranous AQP5 in cells in the presence of chlorpheniramine ⁴¹. Cholinergic stimulation plays an important role in inflammatory airway diseases. It was revealed that methacholine, a parasympathomimetic drug, disrupted airway surface fluid homeostasis and this effect was due to a decrease in AQP5 level after methacholine-induced activation of the NF-kB pathway, while dexamethasone reversed this effect by decreasing ⁴². Many studies have shown that IL-13 is the central regulator of disorders, such as asthma, and some studies have argued that IL-13 blockade prevents allergen-induced airway inflammation 20,43. Skowron-Zwarg et al. found that IL-13 did not affect AQP3 or AQP4 expression but removed AQP5 expression. They suggested that IL-13 can mediate AQP5 inhibition by activating TNF- $\alpha^{20,44}$.

AQPs and Chronic Rhinosinusitis

Chronic rhinosinusitis (CRS) is a common upper respiratory tract disease that occurs due to dysregulation of the inflammatory response, generally due to microbial infection in the nasal cavity and sinus. CRS has been divided into two: CRS with nasal polyps, which tends to Th2 cytokine polarization and CRS without nasal polyps, which is mostly associated with a Th1-type response ^{3,20}. Histologically, nasal polyps are characterized by sparse fibrous cell edematous fluid, few mucus glands without innervation, squamous metaplasia of the surface epithelium, proliferation of stromal and epithelial elements, and allergens. AR arises from type I hypersensitivity reactions associated thickening of the basal membrane 45. It has been put forward that such with immunoglobulin E mediating allergic responses. AR affects immunological and histological differences are associated with proteins, such as AQPs²⁰. In a study comparing nasal polyp tissue and expression of AQPs in the vomeronasal sensory epithelium, it has normal tissue, it was revealed that AQP1 is abundantly expressed in polyp tissue fibroblasts, especially in the subepithelial area, in the periphery of seromucous glands and endothelial cells of venules. It has been suggested that AQP1, which is higher in polyp tissue than normal tissue, is among the mechanisms contributing to tissue edema ⁴⁶. In a study comparing AQP5 expressions in normal control, CRS without nasal polyps and CRS with nasal polyps, it was observed that the epithelial expression of AQP5 was lower in CRS tissues with nasal polyps than in the other two groups. Thus, the authors have argued that the mucosal epithelial barrier is compromised in the context of CRS disease, notably CRS with nasal polyps and that loss of AQP5, which can act as a tight junction protein, plays a role in the occurrence of mucosal edema and the pathophysiology of nasal polyp formation³. In a similar study reporting a decrease in AQP5 in tissues from patients who had CRS with nasal polyps, it was concluded that loss of AQP5 results in edema and polyp formation due to disruption of tight junction regulation of cell volume and failure to maintain epithelial water homeostasis, and the production of dark secretions, which are typical features of CRS with nasal polyps 47. AQP5 expression levels in the control, CRS and dexamethasone treatment groups were compared in a study conducted in rats in which an experimental CRS model was established. Compared with the other two groups, AQP5 expression significantly increased in the dexamethasone-treated group. The findings showed that the infectious agent *Staphylococcus aureus*, which was used in the study to create a CRS model, decreased AQP5 expression by destroying the ciliary epithelium and glandular tissue where AQP5 is primarily localized in the CRS group. It was reported that this tissue destruction was suppressed by dexamethasone and resulted in increased AQP5 expression ⁴⁸. These findings suggest that epithelium and possibly modulate AQP5.

AQPs and Olfactory Epithelium

The olfactory mucosa is a vellowish mucous membrane located in the upper region of the nasal cavity. The olfactory epithelium is made up of olfactory sensory cells, support cells and basal cells. The ends of their dendrites emerge on the epithelial surface and their axons extend to the olfactory bulb in the central nervous system. Mucus protects the olfactory epithelium and allows odors to dissolve so that they can be perceived by olfactory receptor neurons ^{19,20}.

The presence of AQP1, AQP3, AQP4 and AQP5 was determined in a study in which immunohistochemical and immunoblot analyses of the olfactory mucosa of normal rats were conducted. AQP1 expression in olfactory mucosa was found in endothelial cells of blood vessels and surrounding connective tissue cells. On the other hand, AOP1 was not detected in olfactory sensory cells in the olfactory epithelium. AQP3 was abundantly localized to supporting cells and basal cells, whereas AQP4 expression was limited to basal cells. In Bowman's gland, AQP5 was localized to the apical membranes of secreting acinar cells, while expression of AQP3 and AQP4 was detected in the basolateral membrane. A similar localization pattern was detected in the duct cells of Bowman's gland. Drawing on these findings, it has been suggested that there is a different localization pattern for AQPs in the olfactory epithelium and that AQP3, AQP4 and AQP5 in the Bowman gland may play a key role in establishing and maintaining secretory processes that generate a favorable microenvironment for olfactory perception on the apical surface of olfactory dendrites ^{20,34}. In animal experiments, the findings showed that AQP1, AQP3 and AQP4 were expressed in the olfactory mucosa. It has been suggested that AQP4 has a function in olfaction by showing that AQP4 is strongly expressed in the glomerulus, the synaptic unit of the olfactory bulb. Consistent with this finding, a study with AQP4-knockout mice found a reduced sense of smell ^{49,50,51}.

The other chemosensory epithelium with olfactory function in the nasal cavity mucosa is the vomeronasal sensory epithelium. It is anatomically and physiologically distinct from the olfactory system and is a part of the nasal chemosensory system, considered a

differential expression of mucosal water membrane permeability chemosensory organ for pheromones ^{20,52}. In a study investigating the been revealed that AOP1 is localized in blood vessels and is especially abundant in the cavernous tissues of the non-sensory mucosa. AOP5 was detected in the apical membrane of gland acinar cells. AQP3 was detected in basal cells of the non-sensory epithelium. Meanwhile, AQP4 was higher in sensory cells of the sensory epithelium. The findings have shown that AQP4 is specifically localized in the nerve fiber bundles originating from neuronal sensory cells, in the plasma membrane of each axon, and in the lateral membranes of dendrites ⁵² In a study on mice, it has been shown that AQP2, AQP3, AQP4 and AQP5 are expressed at different periods in the olfactory epithelium and vomeronasal sensory epithelium during the embryonic and postnatal periods, depending on the developmental stage. Based on the varying distribution of AQPs by developmental stage, it has been argued that this may be one reason why the olfactory epithelium and vomeronasal sensory epithelium display distinct characteristics as they grow ⁵³. The glia surrounding the olfactory covers or encloses large groups of olfactory receptor neurons along their course towards the olfactory nerve and glomerular layers of the lamina propria and olfactory bulb. AQP1 was expressed in glial cells surrounding the main olfactory bulb but not from olfactory receptor neurons, astrocytes and periglomerular cells. Hence, AQP1 expression is a feature that distinguishes the glial cells surrounding the olfactory system from other central nervous system cell types ⁵⁴.

Conclusion

Expression of AQPs in the sinonasal respiratory epithelium and olfactory epithelium suggested that these proteins may contribute to the pathogenesis of nasal congestion, rhinorrhea, AR and CRS, as well as playing a role in normal physiological processes, such as treating patients with polyps may require strategies that target the humidification of inhaled air and olfactory functions. The expression of AQPs is different in inflammatory diseases, such as AR or CRS than that in normal tissues. Under physiological conditions, the expression of AQPs in the olfactory epithelium differs from that in the respiratory epithelium. In our study, by investigating the relationship between AQPs and sinonasal epithelium, the physiopathological connections between allergic rhinitis, rhinosinusitis, and the olfactory system were revealed through a comprehensive perspective. More comprehensive clinical and experimental findings are needed to fully elucidate the role of aquaporins in the pathophysiology of the sinonasal mucosa. Investigating the regulation and functional roles of AQPs will provide novel insights into the diagnosis and prognosis of facilitate the development of potential therapeutics diseases and for diseases and olfactory epithelium, such as the treatment of AR and CRS.

Conflict of interest

Authors declare that they have no financial interests or personal conflicts that may affect the study in this article.

Financial disclosure

The authors declared that this study has received no financial support.

REFERENCES

4.

5.

- Wang W, Wang X, Ma L, Zhang R. Histamine downregulates aquaporin 5 in human nasal epithelial cells. American journal of rhinology & allergy. 2015;29(3):188-192.
- 2. London NR, Ramanathan M. The role of the sinonasal epithelium in allergic rhinitis. Otolaryngologic Clinics of North America. 2017;50 (6):1043-1050.
- 3. Shikani AH, Sidhaye VK, Basaraba RJ, Shikani HJ, Alqudah MA, Kirk N et al. Mucosal expression of aquaporin 5 and epithelial barrier proteins in chronic rhinosinusitis with and without nasal polyps. American journal of otolaryngology. 2014;35(3):377-383.
 - Chunling L, Wang W. Molecular biology of aquaporins. Aquaporins. $2017 \cdot 1-34$
 - Alp G, Bosgelmez II, Oztas Y. Aquaporins: A Multidisciplinary Perspective on the Water Channel Proteins. Acta Medica. 2020;51(2):30-42.

- Verkman AS. Aquaporins in clinical medicine. Annual review of medicine. 2012;63:303-316
- Day RE, Kitchen P, Owen DS, Bland C, Marshall L, Conner AC et al. 32. Human aquaporins: regulators of transcellular water flow. *Biochimica et Biophysica Acta (BBA)-General Subjects*. 2014;1840(5):1492-1506.
- Azad AK, Raihan T, Ahmed J, Hakim A, Emon TH, Chowdhury PA. 33. Human Aquaporins: Functional Diversity and Potential Roles in Infectious and Non-infectious Diseases. *Frontiers in Genetics*. 2021;12:344.
 34.
- Adeoye A, Odugbemi A, Ajewole T. Structure and Function of Aquaporins: The Membrane Water Channel Proteins. *Biointerface Res. Appl. Chem.* 2021;12:690-705.
- Shchepareva ME, Zakharova MN. Functional role of aquaporins in the nervous system under normal and pathological 35. conditions. *Neurochemical Journal*. 2020;14(1):1-8.
- Verkman AS, Anderson MO, Papadopoulos MC. Aquaporins: important 36. but elusive drug targets. *Nature reviews Drug discovery*. 2014;13(4):259-277.
- 12. Brown PD, Davies SL, Speake T, Millar ID. Molecular mechanisms of 37. cerebrospinal fluid production. *Neuroscience*. 2004;957–970.
- 13. Borsani E. Aquaporins in sensory and pain transmission. Curr. *Neuropharmacol.* 2010;8:122–127.
- Tran TL, Bek T, Holm L, La Cour M, Nielsen S, Prause JU et al. Aquaporins 6-12 in the human eye. *A cta Ophthalmol.* 2013;91:557–563.
- Schey KL, Wang Z, Wenke JL, Qi Y. Aquaporins in the eye: expression, function, and roles in ocular disease. *Biochim. Biophys. Acta.* 2014;1840:1513–1523. doi: 10.1016/j.bbagen.2013.10.037
- Jung SY, Kim SS, Kim YI, Kim SH, Yeo SG. A review: expression of 40. aquaporins in otitis media. *Int. J. Mol. Sci.* 2017;18:2164. doi: 10.3390/ ijms18102164
- Patel R, Kevin Heard L, Chen X, Bollag WB. Aquaporins in the skin. 41. *Adv. Exp. Med. Biol.* 2017;969:173–191. doi: 10.1007/978-94-024-1057-0_11
- Su W, Cao R, Zhang XY, Guan Y. Aquaporins in the kidney: physiology and pathophysiology. *Am. J. Physiol. Renal Physiol.* 2020;318:F193–42. F203
- Jafek BW. Ultrastructure of human nasal mucosa. *The Laryngoscope*. 1983;93(12):1576-1599.
- Jung SY, Park DC, Kim SS, Yeo SG. Expression, distribution and role of 43. aquaporins in various rhinologic conditions. *International Journal of Molecular Sciences*. 2020;21(16):5853.
- Betlejewski S, Betlejewski A. The influence of nasal flow aerodynamics 44. on the nasal physiology. *The Polish Otolaryngology*. 2008;62(3):321-325.
- Nielsen S, King LS, Christensen BM, Agre P. Aquaporins in complex tissues. II. Subcellular distribution in respiratory and glandular tissues of rat. *American Journal of Physiology-Cell Physiology*. 1997;273(5):1549-1561.
- Tsunoda SP, Wiesner B, Lorenz D, Rosenthal W, Pohl P. Aquaporin-1, nothing but a water channel. *Journal of Biological Chemistry*. 2004;279 (12):11364-11367.
- 24. Ablimit A, Matsuzaki T, Tajika Y, Aoki T, Hagiwara H, Takata K. Immunolocalization of water channel aquaporins in the nasal olfactory mucosa. *Archives of histology and cytology*. 2006; 69(1):1-12.
- Seno S, Ogawa T, Shibayama M, Kouzaki H, Shimizu T. Expression and localization of aquaporin 1, 2, 3, 4, and 5 in human nasal mucosa. *American Journal of Rhinology & Allergy*. 2012; 26(3):167-171.
- Frauenfelder C, Woods C, Hussey D, Ooi E, Klebe S, Carney AS. Aquaporin expression profiles in normal sinonasal mucosa and chronic rhinosinusitis. In International Forum of Allergy & Rhinology. 2014;4 49. (11):901-908.
- 27. Fenton RA, Pedersen CN, Moeller HB. New insights into regulated aquaporin-2 function. *Current opinion in nephrology and hypertension*. 2013;22(5):551-558.
- Saadoun S, Papadopoulos MC. Aquaporin-4 in brain and spinal cord oedema. *Neuroscience*. 2010;168:1036–1046.
- Gleiser C, Wagner A, Fallier-Becker P, Wolburg H, Hirt B, Mack AF. (2016). Aquaporin-4 in astroglial cells in the CNS and supporting cells of sensory organs—a comparative perspective. *International journal of* 52. *molecular sciences*. 2016;17(9):1411.
- Song Y, Verkman AS. Aquaporin-5 dependent fluid secretion in airway submucosal glands. *Journal of Biological Chemistry*. 2001;276(44):41288 -41292.
- Matsuzaki T, Suzuki T, Koyama H, Tanaka S, Takata K. Water channel protein AQP3 is present in epithelia exposed to the environment of

possible water loss. *Journal of Histochemistry & Cytochemistry*. 1999;47 (10):1275-1286.

- Kishida K, Kuriyama H, Funahashi T, Shimomura I, Kihara S, Ouchi N et al. Aquaporin adipose, a putative glycerol channel in adipocytes. *Journal of Biological Chemistry*. 2000;275(27):20896-20902.
- Seo YJ, Choi JY. Expression and localization of aquaporin water channels in human middle ear epithelium. *Otology & Neurotology*. 2015;36(7):1284-1289.
- 34. Sakai H, Sato K, Kai Y, Shoji T, Hasegawa S, Nishizaki M et al. Distribution of aquaporin genes and selection of individual reference genes for quantitative real-time RT-PCR analysis in multiple tissues of the mouse. *Canadian journal of physiology and pharmacology*. 2014;92 (9):789-796.
- Chong SN, Chew FT. Epidemiology of allergic rhinitis and associated risk factors in Asia. World Allergy Organization Journal. 2018;11:17.
- Cagnani CEB, Solé D, Díaz SNG, Zernotti ME, Sisul JC, Borges MS et al. Allergic rhinitis update and its impact on asthma (ARIA 2008). *Latin American perspective. Revista Alergia México.* 2009;56(2):56-63.
- Jutel M, Blaser K, Akdis CA. Histamine in allergic inflammation and immune modulation. *International archives of allergy and immunology*. 2005;137(1): 82-92.
- Ma T, Song Y, Gillespie A, Carlson EJ, Epstein CJ, Verkman AS. Defective secretion of saliva in transgenic mice lacking aquaporin-5 water channels. *Journal of Biological Chemistry*. 1999;274(29):20071-20074.
- Xu R, Xu G, Shi J, Wen WA Correlative study of NF-κB activity and cytokines expression in human chronic nasal sinusitis. *The Journal of Laryngology & Otology*. 2007;121(7):644-649.
 - Wang W, Zheng M. Role of cAMP-PKA/CREB pathway in regulation of AQP 5 production in rat nasal epithelium. *Rhinology*. 2011; 49(4):464-469.
- Chang YL, Lin CS, Wang HW, Jian KR, Liu SC. Chlorpheniramine attenuates histamine-mediated aquaporin 5 downregulation in human nasal epithelial cells via suppression of NF-κB activation. *International journal of medical sciences*. 2017; 14(12):1268.
- Chang YL, Jian KR, Lin CS, Wang HW, Liu SC. Dexamethasone attenuates methacholine mediated aquaporin 5 downregulation in human nasal epithelial cells via suppression of NF□κB activation. In International Forum of Allergy & Rhinology. 2018;8(1):64-71.
- Grünig G, Warnock M, Wakil AE, Venkayya R, Brombacher F, Rennick DM et al. Requirement for IL-13 independently of IL-4 in experimental asthma. *Science*. 1998;282: 2261–2263.
- Skowron-Zwarg M, Boland S, Caruso N, Coraux C, Marano F, Tournier F. Interleukin-13 interferes with CFTR and AQP5 expression and localization during human airway epithelial cell diferentiation. *Exp. Cell Res.* 2007;313:2695–2702.
- Pawankar R. Nasal polyposis: an update. *Current opinion in allergy and clinical immunology*. 2003;3(1):1-6.
- 46. Altuntaş A, Yılmaz MD, Aktepe F, Kahveci OK, Derekoy S, Dilek H et al. Expression and distribution of aquaporin-1 in nasal polyps: Does it have any significance in edema formation?. *American journal of rhinology*. 2006;20(1):128-131.
- Pistochini A, Rossi F, Gallo S, Pirrone C, Preti A, Gornati, R et al. Multiple gene expression profiling suggests epithelial dysfunction in polypoid chronic rhinosinusitis. *Acta Otorhinolaryngologica Italica*. 2019;39 (3):169.
- 48. Yu CJ, Cui XY, Lu L, Yang J, Chen B, Zhu CW et al. Effects of glucocorticoid on the expression and regulation of aquaporin 5 in the paranasal sinus of rats with chronic rhinosinusitis. *Experimental and Therapeutic Medicine*. 2017;13(5):1753-1756.
- Solbu TT, Holen T. Aquaporin pathways and mucin secretion of Bowman's glands might protect the olfactory mucosa. *Chemical senses*. 2012;37(1):35-46.
- Sørbø JG, Moe SE, Holen T. Early upregulation in nasal epithelium and strong expression in olfactory bulb glomeruli suggest a role for Aquaporin-4 in olfaction. *FEBS letters*. 2007;581(25):4884-4890.
- Lu DC, Zhang H, Zador Z, Verkman AS. Impaired olfaction in mice lacking aquaporin □4 water channels. *The FASEB Journal.* 2008;22 (9):3216-3223.
- Ablimit A, Aoki T, Matsuzaki T, Suzuki T, Hagiwara H, Takami S, Takata K. Immunolocalization of water channel aquaporins in the vomeronasal organ of the rat: expression of AQP4 in neuronal sensory cells. *Chemical senses*. 2008;33(5):481-488.
- Merigo F, Mucignat Caretta C, Cristofoletti M, Zancanaro C. Epithelial membrane transporters expression in the developing to adult mouse vomeronasal organ and olfactory mucosa. *Developmental neurobiology*.

2011;71(10):854-869.
54. Shields SD, Moore KD, Phelps PE, Basbaum AI. Olfactory ensheathing glia express aquaporin 1. *Journal of Comparative Neurology*. 2010;518 (21):4329-4341.